

Supplementary information for:

CAF-1 deposits newly synthesized histones during DNA replication using distinct mechanisms on the leading and lagging strands

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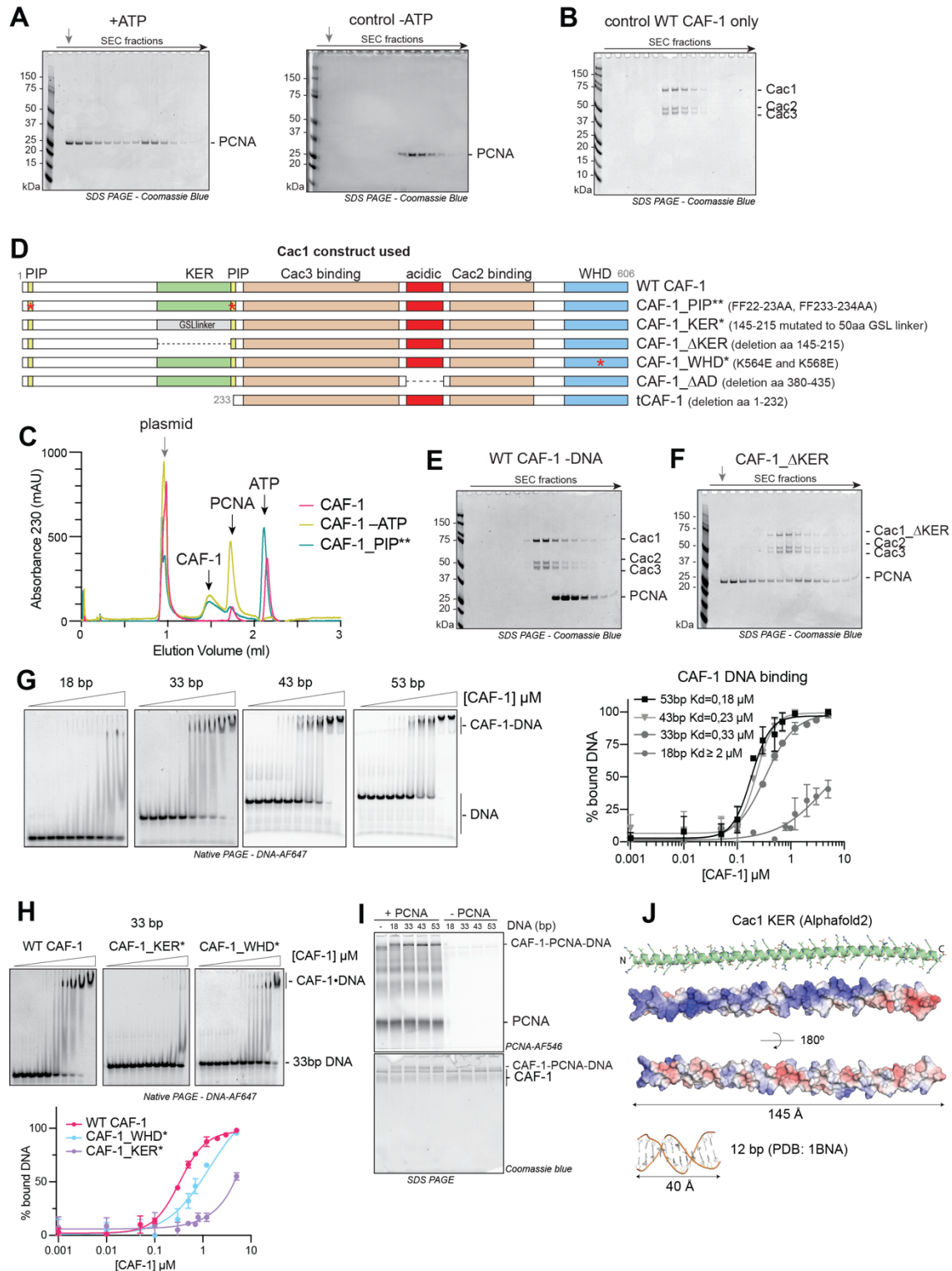
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Supplemental Figure S1

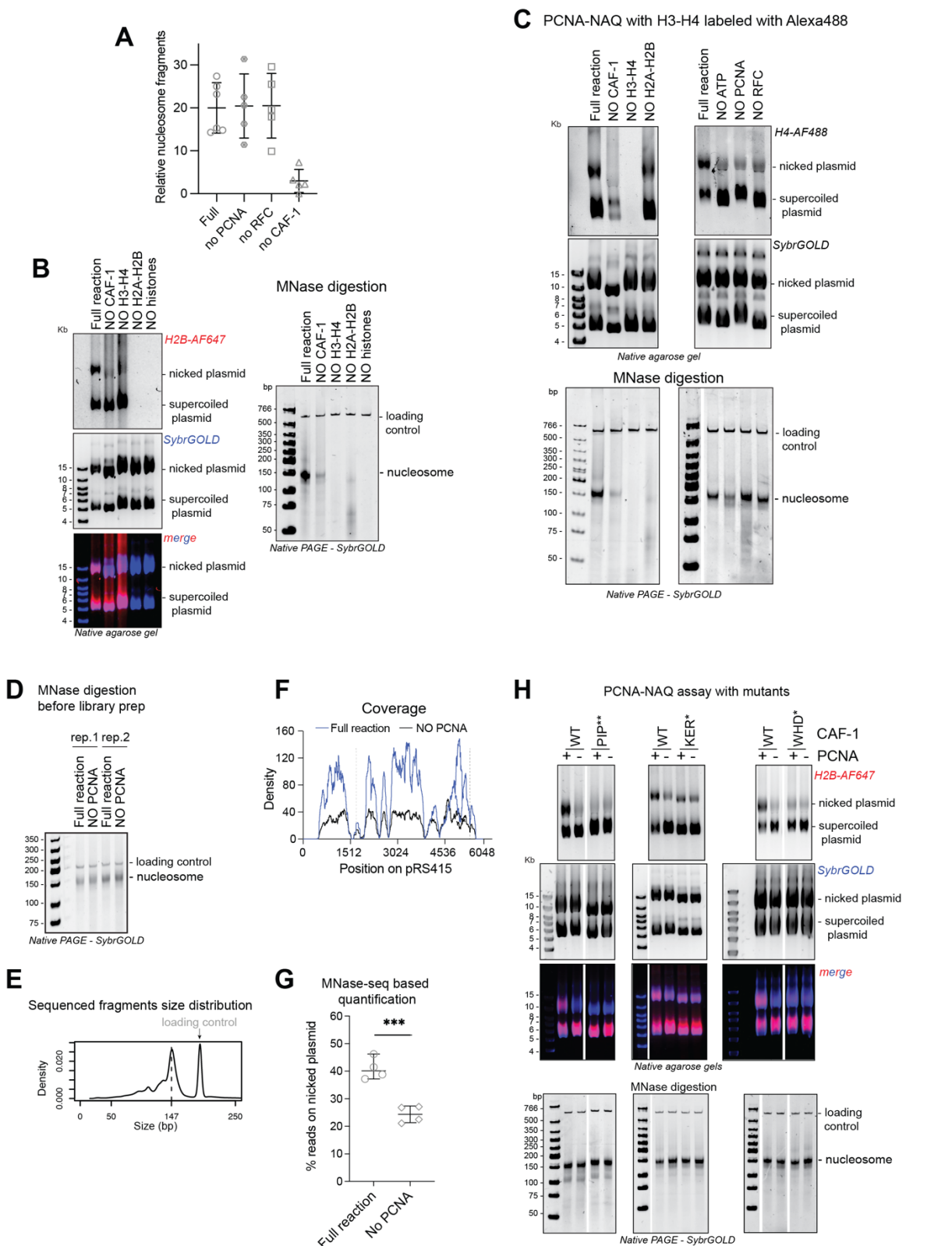


Supplementary Figure S1

A) SDS PAGE following separation on SEC of a PCNA loading reaction on DNA plasmids (on the left) and the control omitting ATP (on the right). The grey arrow indicates the elution volume of the plasmid DNA. **B)** SDS PAGE following separation on SEC of WT CAF-1 only. **C)** Chromatogram (230 nm signal) of the Superose 6 runs whose SDS PAGE gels are shown in Figure 1A-C. The grey arrow indicates the elution volume of the plasmid DNA.

Elution volume for CAF-1, PCNA and ATP are marked by black arrows. **D)** Cartoons of the Cac1 (large CAF-1 subunit) domains and the mutants used in this study. All complexes contain Cac2 and Cac3, in addition to the indicated Cac1 construct. **E)** SDS PAGE following separation on SEC of a control reaction where the DNA plasmid was omitted, from a CAF-1-PCNA binding experiment as in Figure 1A. **F)** SDS PAGE following separation on SEC of a CAF-1-PCNA binding reaction on nicked DNA plasmid using a CAF-1_ΔKER mutant. The grey arrow indicates the elution volume of the plasmid DNA. **G)** EMSA experiments and quantification of CAF-1 binding to double-stranded DNA fragments. Each 18-33-43-53 bp DNA fragment carries a AF647 fluorophore for detection and quantification. Disappearance of the unbound DNA band was used to calculate binding at different CAF-1 concentrations. Binding affinities K_d result from fitting the calculate data in GraphPad. 18 bp data is fitted with a One Site total binding curve, while the data with 33-43-53 bp DNA was fit accounting for cooperativity. The Hill coefficients obtained for these curves are 1.6, 2.7, 2.4 respectively. Means \pm SD is shown for each data point. At least three replicates were done for each experiment. **H)** EMSA experiments and quantification of binding to a 33-bp double-stranded DNA fragment of CAF-1_KER* and CAF-1_WHD* mutants. Disappearance of the unbound DNA band was used to calculate binding at different CAF-1 concentrations. Means \pm SD is shown for each data point. At least three replicates were done for each experiment. **I)** Crosslinking experiment between CAF-1 (3 μ M) and labeled PCNA (4.5 μ M) on DNA fragments (1.5 μ M) of various sizes. DNA was not digested in these reactions. RFC and ATP were not added to actively load PCNA. These are the full gels of Figure 1G. **J)** Alphafold model of the KER domain in Cac1 (residues 128-226). Cartoon with sticks and electrostatics are shown. The electrostatics are calculated with the APBS plugin in Pymol. Blue is +5kEV and Red is -5kEV. A 12bp structure of B-DNA (PDB: 1BNA) is shown at the same scale of the KER domain for length comparison.

Supplemental Figure S2



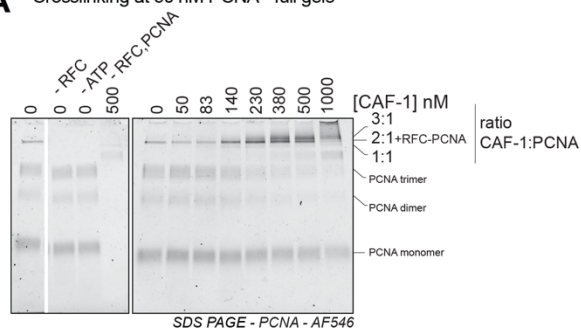
Supplementary Figure S2

A) Bioanalyzer-based quantification of nucleosome assembly (140-160 bp fragments) relative to the 621 bp loading control shows that CAF-1 is the main nucleosome assembly factor in our reactions. **B)** Left panel: Native agarose gel of control PCNA-NAQ assay conditions. Fluorescence signal for H2B-T112C labeled with AF647 (H2B-AF647) or DNA (SybrGOLD), and their overlay are shown. H2B fluorescence on the nicked plasmid (top panel) represents PCNA-dependent histone deposition. Right panel: Native PAGE stained with SybrGOLD of protected

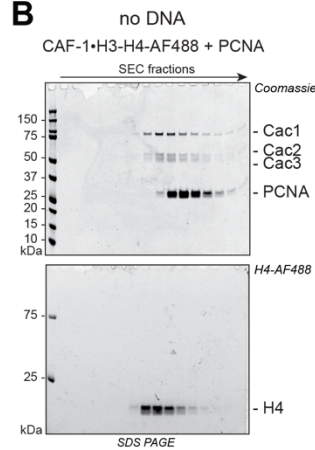
DNA fragments following MNase digestion. 150bp DNA fragments are characteristic of nucleosomal DNA, a 621bp loading control is used to monitor DNA retrieval during the purification procedure. **C)** Top panels: Native agarose gel of control PCNA-NAQ assay conditions where instead of fluorescently labeled H2A-H2B, we used labeled H3-H4. Fluorescence signal for H4-E63C labeled with AF488 (H4-AF488) is shown. Subsequently we stained with SybrGOLD to image DNA. H4 fluorescence on the nicked plasmid (top panel) represent PCNA-dependent histone deposition. Bottom panels: Native PAGE stained with SybrGOLD of protected DNA fragments following MNase digestion. 150bp DNA fragments are characteristic of nucleosomal DNA, a 621bp loading control is used to monitor DNA retrieval during the purification procedure. **D)** MNase digestion gel of PCNA-NAQ assay samples obtained for NGS analysis. These reactions contained a nicked and supercoiled plasmid with different sequences (pRS415 or pLox3). We used a 207 bp DNA loading control containing a 601-widom sequence. **E)** Fragment size distribution of the sequenced reads confirms the dominance of a 150 bp size after MNase digestion and sequencing. **F)** Example of reads coverage on the nicked plasmid when pRS415 was used in the full reaction or in a negative control reaction omitting PCNA. Dashed lines show the sites of nicking. **G)** Quantification of the PCNA-dependent nucleosome assembly activity based on the NGS reads of WT CAF-1 and a no-PCNA control reaction. The percentage of reads on the nicked plasmid is shown over the total number of reads (for both plasmids). Means \pm SD is shown, and an unpaired t-test was applied to determine statistical significance, *** $p > 0,001$. **H)** Native agarose gel (top) of control PCNA-NAQ assay conditions with CAF-1 mutants. Fluorescence signal for H2B-T112C labeled with AF647 (H2B-AF647) or DNA (SybrGOLD), and their overlay are shown. H2B fluorescence on the nicked plasmid (top panel) represents PCNA-dependent histone deposition. Bottom: Native PAGE stained with SybrGOLD of protected DNA fragments following MNase digestion. 150bp DNA fragments are characteristic of nucleosomal DNA, a 621bp loading control is used to monitor DNA retrieval during the purification procedure. Quantifications of H2B fluorescence are shown in Figure 2C.

Supplemental Figure S3

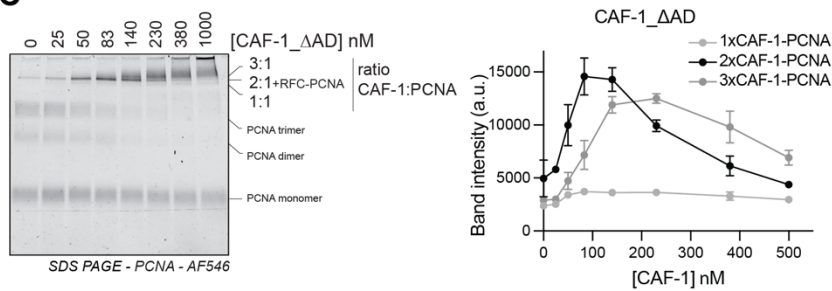
A Crosslinking at 50 nM PCNA - full gels



B



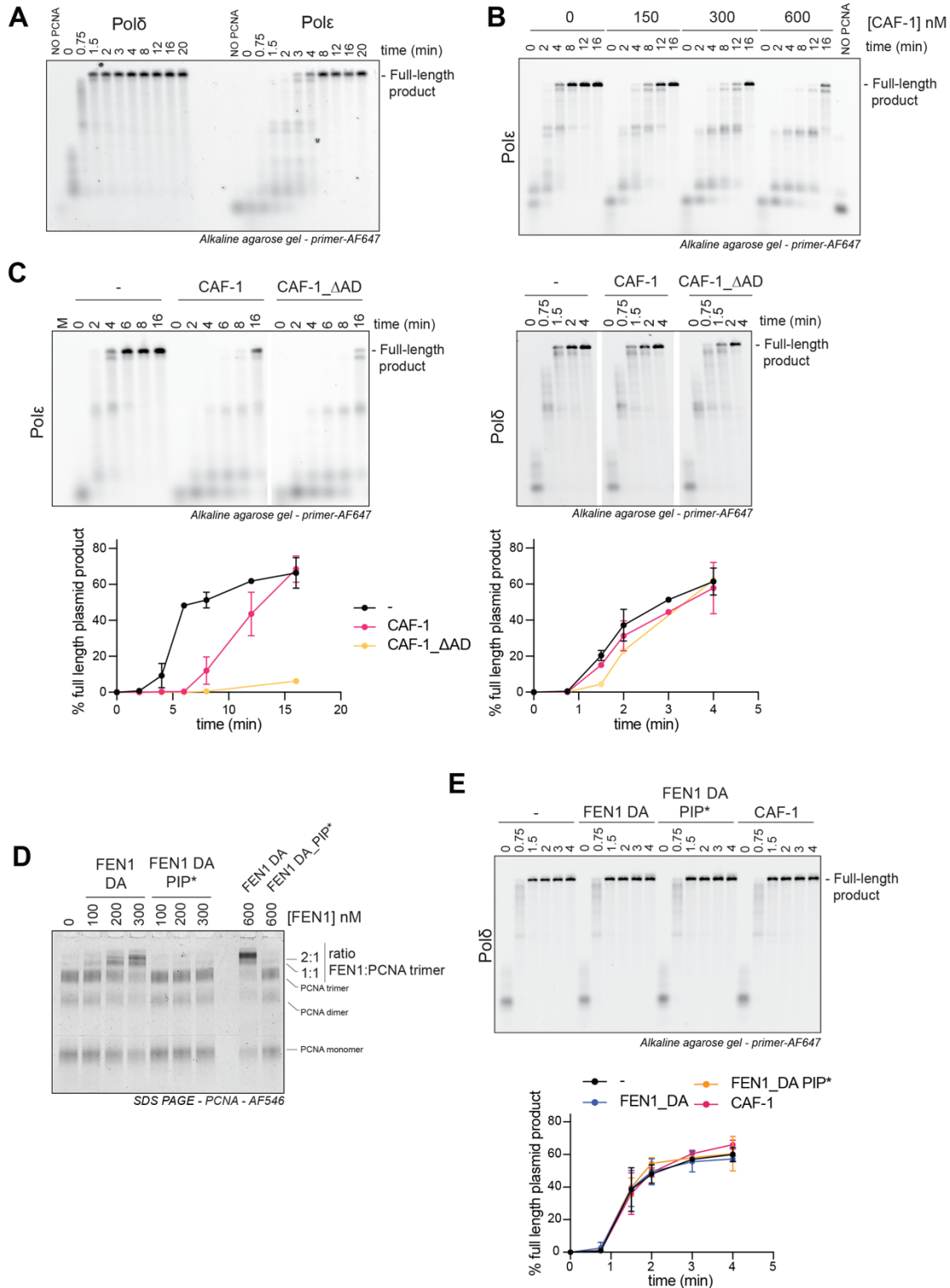
C



Supplementary Figure S3

A) Full gels of data in Figure 3C. PCNA fluorescence scan of SDS-PAGE of crosslinking experiment after DNA digestion, of reactions containing 50 nM PCNA, 15 nM RFC, 15 nM pUC19 and increasing CAF-1 concentrations. **B)** SDS PAGE after SEC of a reaction containing PCNA (30 μ M), and CAF-1 preloaded with labeled H3-H4 (5 μ M). No DNA or RFC is present in these reactions. After SDS PAGE run, we scan the gel for the H4-488 nM fluorescence (bottom), followed by staining with Coomassie (top). **C)** PCNA fluorescence scan of SDS-PAGE following protein-protein crosslinking after DNA digestion. These reactions contain 50 nM PCNA, 15 nM RFC, 15 nM pUC19 and increasing CAF-1_ΔAD concentrations. On the right: quantification of the CAF-1-PCNA bands. Mean \pm SD is shown of three replicates.

Supplemental Figure S4

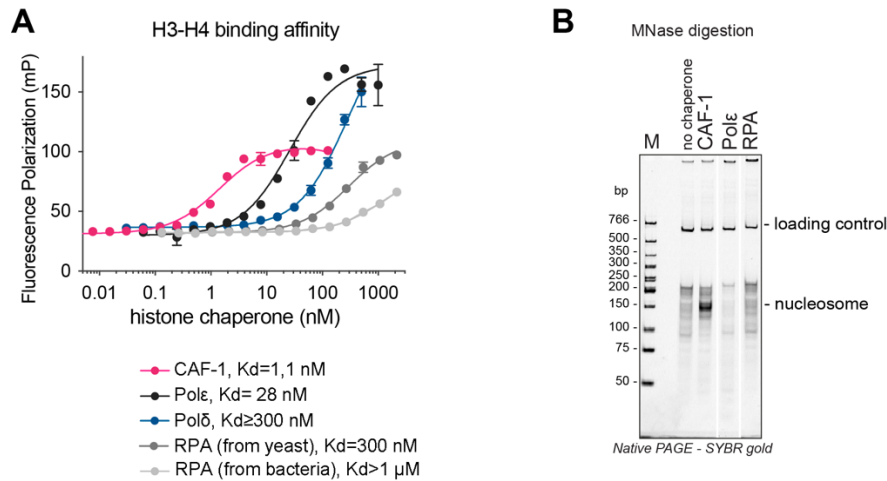


Supplementary Figure S4

A) Fluorescence (primer signal) scan of denaturing alkaline agarose gel of primer extension reactions with Polδ or Polε. The polymerases were at 120 nM, PCNA 480 nM. Both polymerases are active with different kinetics. Both polymerases depend on PCNA for activity. The NO-PCNA control lanes contain reactions that incubated for 20 minutes. **B)** Fluorescence (primer signal) scan of denaturing alkaline agarose gel of primer extension reactions with Polε in the presence of increasing amounts of WT CAF-1 (150-300-600 nM). **C)** Fluorescence (primer signal) scan of denaturing alkaline agarose gel of primer extension reactions with Polε (left) or Polδ (right) in the presence of CAF-1 WT or CAF-1_ΔAD (300 nM). Bottom: Quantification of the full-length product band relative to the total fluorescence in each lane (expressed as percentages). Mean ±SD are shown for three replicates. **D)**

PCNA fluorescence scan of SDS-PAGE following protein-protein crosslinking. These reactions contain 50 nM PCNA and increasing FEN1_DA or FEN1_DA PIP* concentrations. **E**) Left panel: fluorescence (primer signal) scan of denaturing alkaline agarose gel of primer extension reactions with Pol δ in the presence of WT CAF-1, FEN1_DA and FEN1_DA PIP* (all at 300 nM). Right panel: Quantification of the full-length product band relative to the total fluorescence in each lane (expressed as percentages). Mean \pm SD are shown for three replicates.

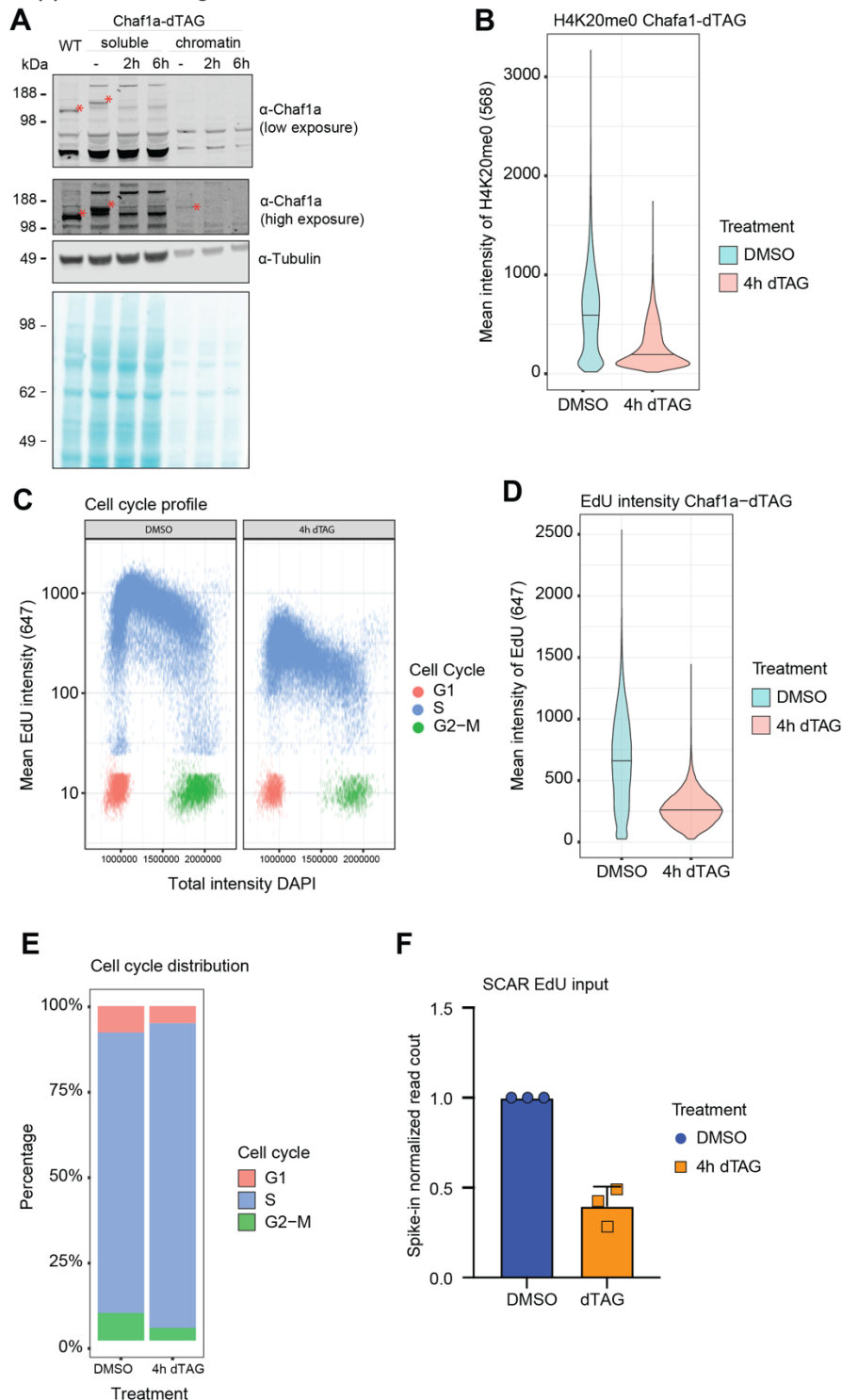
Supplemental Figure S5



Supplementary Figure S5

A) Fluorescence polarization experiment to test binding to H3-H4 (where H4 is labeled at E63C using AF488). CAF-1, Pol ϵ , Pol δ , RPA expressed in yeast (used in Figure 7A) and RPA expressed in bacteria (used in Figure 4-5) were titrated to a solution containing 10 nM of labeled H3-H4. The data points were fit to a one site binding curve and the K_d were calculated in GraphPad. **B)** MNase digestion of a NAQ reaction where each histone chaperone was incubated with the histone octamer and subsequently with 207 bp DNA. Bands at 150 bp represent nucleosomes and these are observed only in the presence of CAF-1. RPA produced from yeast cells was used. A 621 bp loading control DNA is used to control for sample retrieval during the DNA purification step.

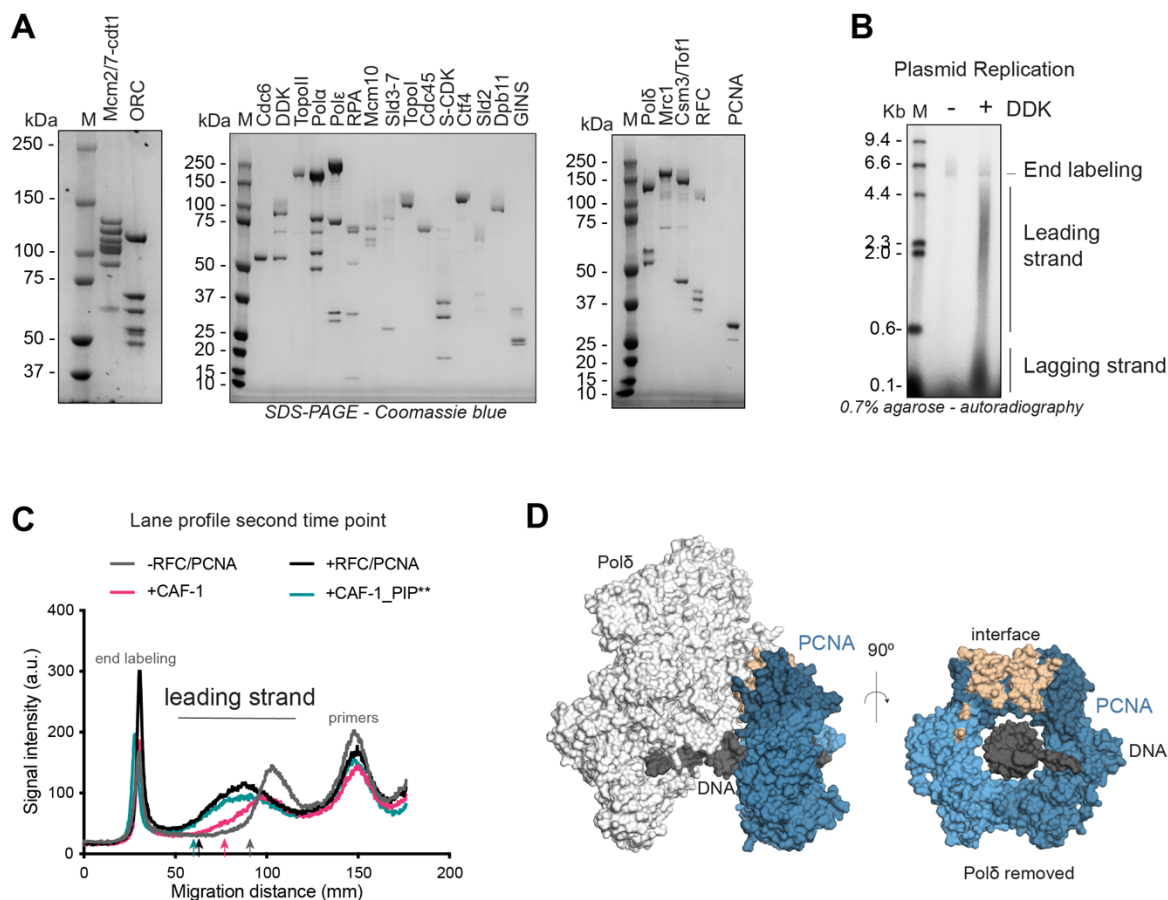
Supplemental Figure S6



Supplementary Figure S6

A) Western blot analysis of soluble and chromatin fractions of mES cells treated with DMSO and dTAG for the indicated times. A WT cell line is shown as a control. **B)** Immunofluorescence results of mean H4K20me0 intensities in mESC upon CAF-1 depletion with dTAG shows a decrease in H4K20me0. **C)** Immunofluorescence results of mean EdU intensity vs total DAPI intensity. **D)** Mean EdU intensity in DMSO vs dTAG treated cells. **E)** Cell cycle distribution based on mean EdU intensity and total EdU intensity. Immunofluorescent data are represented from two replicates. **F)** Spike-in normalized input reads shows decreased EdU incorporation after depletion of CAF-1.

Supplemental Figure S7



Supplementary Figure S7

A) SDS PAGE of protein preps that were used to reconstitute the yeast replisome. **B)** Autoradiography scan of denaturing agarose gel separation using DNA replication products, from an end-point plasmid replication experiment containing all yeast replisome components or, as control, omitting DDK. **C)** Example of lane profiles during pulse-chase experiment after 5 minutes of addition of the chase, as obtained by ImageQuant. The arrows indicate the front of the leading strand (created automatically by ImageQuant), used as the max size of replicated products to calculate the max replication rates. **D)** Surface visualization of yeast Pol δ (in white) bound to PCNA (blue) on DNA (dark gray) from PDB 7KC0. In what we show the PCNA residues involved in the Pol δ interaction. On the right panel Pol δ is not displayed.

Supplementary Information

Supplementary Table S1: Primers used for EMSA.

DNA primers	Reference
DNA oligo 18mer forward: GTCTACGAGCAATTGAGC	Mattiroli et al., 2017
DNA oligo 18mer reverse: GCTCAATTGCTCGTAGAC	Mattiroli et al., 2017
DNA oligo 33mer forward: GCTGTCTACGAGCAATTGAGCGGCCTCGGCACC	Mattiroli et al., 2017
DNA oligo 33mer reverse: GGTGCCGAGGCCGCTCAATTGCTCGTAGACAGC	Mattiroli et al., 2017
DNA oligo 43mer forward, 5' conjugated to AlexaFluor 647: CTAGAGCTGTCTACGAGCAATTGAGCGGCCTCGGCACCGGAT	This paper
DNA oligo 43mer reverse: ATCCCGGTGCCGAGGCCGCTCAATTGCTCGTAGACAGCTCTAG	This paper
DNA oligo 53mer forward, 5' conjugated to AlexaFluor 647: CGGTGCTAGAGCTGTCTACGAGCAATTGAGCGGCCTCGGCACCGGGATTCTGA	This paper
DNA oligo 53mer reverse: TCAGAATCCCGGTGCCGAGGCCGCTCAATTGCTCGTAGACAGCTCTAGCACCG	This paper

Supplementary Table S2, DNA sequences for plasmids and linear fragments used in MNase-seq

Name	DNA sequence
pLox3	<p>AACGACCTACACCGAACTGAGATACCTACAGCGTGAGCTATGAGAAAAGCGCCACGCTTCCCGAAGGG AGAAAGGCGGACAGGTATCCGGTAAGCGGCAGGGTCGGAACAGGAGAGCGCACGAGGGGAGCTTCC AGGGGGAAACGCCTGGTATCTTTATAGTCCTGTCGGGTTTTCGCCACCTCTGACTTGAGCGTCGATTTTT GTGATGCTCGTCAGGGGGGCGGAGCCTATGGAAAAACGCCAGCAACGCGGCCTTTTTACGGTTCCTG GCCTTTTGCTGGCCTTTTGCTCACATGTTCTTCTGCGTTATCCCCTGATTGACTTGGGTGCTCTTCT GTGGATGCGCAGATGCCCTGCGTAAGCGGGTGTGGGCGGACAATAAAGTCTTAACTGAACAAAATA GATCTAAACTATGACAATAAAGTCTTAACTAGACAGAATAGTTGTAAGTCAAATCAGTCCAGTTATG CTGTGAAAAAGCATACTGGACTTTTTGTTATGGCTAAAGCAAACCTTTCATTTTCTGAAGTGCAAATTGC CCGTCGTATTAAGAGGGGGCGTGGCCAAGGGCATGTAAAGACTATATTCGCGGCGTTGTGACAATTTA CCGAACAACCTCCGCGGCCGGGAAGCCGATCTCGGCTTGAACGAATTGTTAGGTGGCGGTACTTGGGT CGATATCAAAGTGCATCACTTCTCCCGTATGCCCACTTTGTATAGAGAGCCACTGCGGGATCGTCAC CGTAATCTGCTTGACGTAGATCACATAAGCACCAAGCGCGTTGGCCTCATGCTTGAGGAGATTGATG AGCGCGGTGGCAATGCCCTGCCTCCGGTGTCTCGCCGAGACTGCGAGATCATAGATATAGATCTCACT ACGCGGCTGCTCAAACCTTGGCAGAACGTAAGCCGCGAGAGCGCCAACAACCGCTTCTTGGTGAAG GCAGCAAGCGCGATGAATGTCTTACTACGGAGCAAGTTCCCGAGTAATCGGAGTCCGGCTGATGTT GGGAGTAGGTGGCTACGTCTCCGAACCTCACGACCGAAAAGATCAAGAGCAGCCCGCATGGATTTGAC TTGGTCAGGGCCGAGCCTACATGTGCGAATGATGCCATACTTGAGCCACCTAACTTTGTTTTAGGGC GACTGCCCTGCTGCGTAACATCGTTGCTGCTGCGTAACATCGTTGCTGCTCCATAACATCAAACATCGA CCCACGGCGTAACGCGCTTGTGCTTGGATGCCGAGGCATAGACTGTACAAAAAACAGTCATAACA AGCCATGAAAACCGCCACTGCGCGTACCACCGCTGCGTTCGGTCAAGTTCTGGACCAGTTGCGTG AGCGCATACTACTTGCATTACAGTTTACGAACCGAACAGGCTTATGTCAACTGGGTTCTGTCCTTCA TCCGTTTCCACGGTGTGCGTCACCCGGCAACCTTGGGCAGCAGCGAAGTCGCCATAACTTCGTATAGC ATACATTATACGAAGTTATCTGCCAGGCACATGGGTTTTACTAGTATCGATTGCGGACCTACTCCGGAA TATTAATAGATCATGGAGATAATTAATGATAACCATCTCGCAAATAAATAAGTATTTTACTGTTTTCG TAACAGTTTTGTAATAAAAAAACCTATAAATATCCGGATTATTCATACCGTCCACCATCGGGCGCGG ATCCCGACCATGCATCACCATCACCATCACCATAATCAGTGCGCGAAGGACGCGCGGATCCCGACCAT GCATCACCATCACCATCACCATAATCAGTGCGCGAAGGACATAACTCATGAAGCCTCCAGTATACCCAT CGATTTGCAAGAAAGATACTGCACTGGAAGAAAAACCTAAACTACTTTATGATTACCTAAACACGA ATTCAACAAAGTGGCCGTCCTAACGTGCCAGTTCTTTCCTGATTTAGATACCACTTCGGATGAGCATC GCATCTTGTATCCTCATTTACATCTTCCAAAAACCTGAAGATGAGACCATATATATTAGCAAAATATC CACGTTGGGTCATATAAAATGGTCATCTTTAATAATTTGACATGGACGAAATGGAATTCAAACCGG AGAACTCGACAAGGTTTCCCTCAAACACTTAGTAAATGACATCAGTATTTTCTTCCAAACGGGGAAT GCAATAGGGCAAGATATTTGCCTCAAATCCAGATATTATAGCCGGCGCCTTTCAGATGGTGAATCT ACATATTCGATAGAACAAAAACGCGCTCTACTAGAATAAGACAGTCCAAAAATTCACATCCCTTTGAGA CAAAGCTGTTTGGTTCACATGGTGTATTCAAGACGTGGAGGCAATGGATACTTCTTCGGCAGATATA AATGAGGCGACTTCTTAGCCTGGAACCTGACGACGAGGAGCCCTTTACTTTCTTCTACTCCAACGGC CAAGTTCAAGTTTGGGACATTAACAATATTTCGCATGAGAACCCTATAATAGATTTACCTTAGTGCA ATAAACAGCGACGGAACAGCGGTGAATGATGTAACCTGGATGCCAACACACGATTCCCTCTTGTGCTG TTGTAAGGAAATGCGGTCTCCCTATTAGATCTGAGGACTAAGAAAGAGAAGCTCCAGAGTAACC GTGAAAAACAGATGGTGGAGTAAACTCCTGTAGATTTAACTATAAGAACTCTTAAATCTAGCATCTG CAGATTCAAATGGGAGGCTAAATTTATGGGATATTAGAAACATGAACAAAAGCCCAATCGTACCATG GAGCACGGTACTTCCGTTTCAACTTTAGAATGGAGTCCAAATTTGATACTGTATTGGCAACGGCTGGC CAAGAAGATGGGTTAGTCAAGCTATGGGATACCTCCTGCGAAGAACTATATTTACCATGGTGGTCA TATGCTCGGTGTGAACGACATTTCTGTTGGGACGCTCATGACCCTTGGTTAATGTGCAGTGTGGCAAATG ATAATTCAGTTCACATATGGAAACCTGCAGGAAACCTTGTGGACATTCGTGAGCTCTAGAGCCTGCA GTCTCGACAAGCTTGTGAGAAGTACTAGAGGATCATAATCAGCCATACCACATTTGTAGAGGTTTTAC TTGCTTTAAAAAACCTCCACACCTCCCCCTGAACCTGAAACATAAAAATGAATGCAATTGTTGTTAA CTTGTATTGTCAGCTTATAATGGTTACAAATAAAGCAATAGCATCACAATTTCAAAATAAAGCATT TTTTCACTGCATTCTAGTTGTGGTTTGTCCAACTCATCAATGTATCTTATCATGTCTGGATCTGATCACT GCTTGAGCCTAGAAGATCCGGCTGCTAACAAAAGCCGAAAGGAAGCTGAGTTGGCTGCTGCCACCGC TGAGCAATAACTATCATAACCCCTAGGTGCCATTTTATTACCTCTTCTCCGCACCCGACATAGATCTGG GCCAACTTTTGGCGAAAATGAGACGTTGATCGGCACGTAAGAGGGTCCAACTTACCATAATGAAAT AAGATCACTACCGGGCGTATTTTTGAGTTATCGAGATTTTCAGGAGCTAAGGAAGCTAAAAATGGAGA</p>

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ATGATTTTCTGTTACACCTAACTTTTTGTGTGGTGCCTCCTCTTGTCAATATTAATGTTAAAGTGCAAT
TCTTTTCTTATCACGTTGAGCCATTAGTATCAATTTGCTTACCTGTATTCTTTACATCCTCCTTTTTCT
CCTTCTGTAAATGTATGTAGATTGCGTATATAGTTTCGTCTACCCTATGAACATATTCATTTTGTAAAT
TTCGTGTCGTTTCTATTATGAATTTCAATTTATAAAGTTTATGTACAAATATCATAAAAAAAGAGAATCTT
TTAAGCAAGGATTTCTTAACTTCTTCGGCGACAGCATCACCGACTTCGGTGGTACTGTTGGAACCAC
CTAAATCACCAGTTCTGATACCTGCATCCAAAACCTTTTTAACTGCATCTTCAATGGCCTTACCTCTTCA
GGCAAGTTCAATGACAATTTCAACATCATTGCAGCAGACAAGATAGTGGCGATAGGGTTGACCTTATT
CTTTGGCAAATCTGGAGCAGAACCGTGGCATGGTTCGTACAAACCAATGCCGTGTTCTTGTCTGGCA
AAGAGGCCAAGGACGCAGATGGCAACAAACCAAGGAACCTGGGATAACGGAGGCTTCATCGGAGA
TGATATCACCACATGTTGCTGGTGATTATAATACCATTTAGGTGGGTTGGGTTCTTAACTAGGATCA
TGCGCGCAGAATCAATCAATTGATGTTGAACCTTCAATGTAGGGAATTCGTTCTTGATGGTTTCTCCA
CAGTTTTTCTCCATAATCTTGAAGAGGCCAAAACATTAGCTTTATCCAAGGACCAAATAGGCAATGGTG
GCTCATGTTGTAGGGCCATGAAAAGCGGCCATTCTTGTGATTCTTTGCACTTCTGGAACGGTGTATTGTT
CACTATCCCAAGCGACACCATCACCATCGTCTTCTTTCTTACCAAAGTAAATACCTCCCCTAATTCT
CTGACAACAACGAAGTCAGTACCTTTAGCAAATTGTGGCTTGATTGGAGATAAGTCTAAAAGAGAGTC
GGATGCAAAGTTACATGGTCTTAAAGTTGGCGTACAATTGAAGTCTTTACGGATTTTTAGTAAACCTTG
TTCAGGTCTAACACTACCGGTACCCATTTAGGACCACCCACAGCACCTAACAAAACGGCATCAGCCTT
CTTGAGGGCTTCCAGCGCCTCATCTGGAAGTGGAACACCTGTAGCATCGATAGCAGCACCACCAATTA
AATGATTTTCAAATCGAATTGACATTGGAACGAACATCAGAAATAGCTTTAAGAACCTTAATGGCTT
CGGCTGTGATTTCTGACCAACGTGGTACCTGGCAAAACGACGATCTTCTTAGGGGCGACATAGGG
GCAGACATTAGAATGGTATATCCTTGAATATATATATATATATTGCTGAAATGTA AAAAGGTAAGAAA
AGTTAGAAAAGTAAGACGATTGCTAACACCTATTGGAAAAACAATAGGTCCTTAAATAATATTGTCA
ACTTCAAGTATTGTGATGCAAGCATTAGTCATGAACGCTTCTCTATTCTATATGAAAAGCCGGTCCG
GCGCTCACCTTTCTTTTTCTCCAATTTTTAGTTGAAAAAGGTATATGCGTCAGGCGACCTCTGAA
ATTAACAAAAAATTTCCAGTCATCGAATTTGATTCTGTGCGATAGCGCCCTGTGTGTTCTCGTTATGTT
GAGGAAAAAATAATGGTTGCTAAGAGATTGCAACTCTTGATCTTACGATACCTGAGTATCCACA
GTTAACTGCGGTCAAGATATTTCTTGAATCAGGCGCCTTAGACCGCTCGGCCAAAACAACCAATFACTTG
TTGAGAAAATAGAGTATAATTATCCTATAAATATAACGTTTTTGAACACACATGAACAAGGAAGTACAG
GACAATTGATTTTGAAGAGAATGTGGATTTTGTGTAATTGTTGGGATTCCATTTTTAATAAGGCAATA
ATATTAGGTATATGGATATACTAGAAGTCTCCTCGACCGGTGATATGCGGTGTGAAATACCGCACA
GATGCGTAAGGAGAAAATACCGCATCAGGAAATGTAAGCGTTAATTTTTGTTAAAATTCGCGTTAA
ATTTTTGTTAAATCAGCTCATTTTTTAAACCAATAGGCCGAAATCGGCAAAATCCCTTATAAATCAAAGA
ATAGACCGAGATAGGGTTGAGTGTGTTCCAGTTTGGAAACAAGAGTCCACTATTAAGAACGTGGACT
CCAACGTCAAAGGGCGAAAACCGTCTATCAGGGCGATGGCCCACTACGTGAACCATCACCTAATCA

	AGTTTTTTGGGGTCGAGGTGCCGTAAGCACTAAATCGGAACCTAAAGGGAGCCCCGATTTAGAGC TTGACGGGGAAAGCCGGCGAACGTGGCGAGAAAGGAAGGGAAGAAAGCGAAAGGAGCGGGCGCTA GGGCGCTGGCAAGTGTAGCGGTCACGCTGCGCGTAACCACCACACCCGCCGCGTTAATGCGCCGCT ACAGGGCGCGTCCATTCGCCATTCAGGCTGCGCAACTGTTGGGAAGGGCGATCGGTGCGGGCCTCTT CGCTATTACGCCAGCTGGCGAAAGGGGGATGTGCTGCAAGGCGATTAAGTTGGGTAACGCCAGGGTT TTCCAGTCACGACGTTGTAAAACGACGGCCAGTGAGCGCGCTAATACGACTCACTATAGGGCGAAT TGGGTACCGGGCCCCCCTCGAGGTGACGGTATCGATAAGCTTGATATCGAATTCCTGCAGCCCCGG GGATCCACTAGTTCTAGAGCGGCCGCCACCGCGGTGGAGCTCCAGCTTTTGTCCCTTAGTGAGGGT TAATTGCGCGCTTGGCGTAATCATGGTCATAGCTGTTTCTGTGTGAAATTGTTATCCGCTCACAATTCC ACACAACATACGAGCCGGAAGCATAAAGTGTAAGCCTGGGGTGCCTAATGAGTGAGCTAACTCACA TTAATTGCGTTGCGCTCACTGCCGCTTTCAGTCGGGAAACCTGTCGTGCCAGCTGCATTAATGAATC GGCCAACGCGCGGGGAGAGGGCGGTTTTCGATTGGGCGCTTTCGCTTCCGCTCCTCGCTCACTGACTCGCT GCGCTCGGTGTTTCGGCTGCGGCGAGCGGTATCAGCTCACTCAAAGGCGGTAATACGGTTATCCACAG AATCAGGGGATAACGCAGGAAAGAACATGTGAGCAAAGGCCAGCAAAGGCCAGGAACCGTAAAA AGGCCGCGTTGCTGGCGTTTTTCCATAGGCTCCGCCCCCTGACGAGCATCACAAAATCGACGCTCA AGTCAGAGGTGGCGAAACCCGACAGGACTATAAAGATACCAGGCGTTTCCCCTGGAAGCTCCCTCGT GCGCTCCTGTTCCGACCCTGCCGTTACCGGATACCTGTCCGCTTCTCCCTTCGGGAAGCGTGGC GCTTTCATAGCTCACGCTGTAGGTATCTCAGTTCGGTGTAGGTCGTT
207bp loading control	ATCTAGTATTAATTAATATGAATTCGGATCCACATGCACAGGATGTATATATCTGACACGTGCCTGGAG ACTAGGGAGTAATCCCCTTGGCGGTTAAAACGCGGGGACAGCGCGTACGTGCGTTAAGCGGTGCT AGAGCTGTCTACGACCAATTGAGCGGCCTCGGCACCGGGATTCTCAGGGCGGCCGCGTATAGGGTC CGAT

Supplementary Table S3: - adapters used during MNase-seq (PCNA-NAQ assay)

	Sequence
A1_AACACCTA_FWD	ACACTCTTCCCTACACGACGCTCTTCCGATCTAACACCTA*T
A1_AACACCTA_REV	/5Phos/TAGGTGTTAGATCGGAAGAGCGGTTTCAGCAGGAATGCCGAG
A2_ACGTAGCT_FWD	ACACTCTTCCCTACACGACGCTCTTCCGATCTACGTAGCT*T
A2_ACGTAGCT_REV	/5Phos/AGCTACGTAGATCGGAAGAGCGGTTTCAGCAGGAATGCCGAG
A3_ATATAGGA_FWD	ACACTCTTCCCTACACGACGCTCTTCCGATCTATATAGGA*T
A3_ATATAGGA_REV	/5Phos/TCCTATATAGATCGGAAGAGCGGTTTCAGCAGGAATGCCGAG
A4_CACAGTTG_FWD	ACACTCTTCCCTACACGACGCTCTTCCGATCTCACAGTTG*T
A4_CACAGTTG_REV	/5Phos/CAACTGTGAGATCGGAAGAGCGGTTTCAGCAGGAATGCCGAG
A5_CCTACAAC_FWD	ACACTCTTCCCTACACGACGCTCTTCCGATCTCCTACAAC*T
A5_CCTACAAC_REV	/5Phos/GTTGTAGGAGATCGGAAGAGCGGTTTCAGCAGGAATGCCGAG
A6_CGTCGGCT_FWD	ACACTCTTCCCTACACGACGCTCTTCCGATCTCGTCGGCT*T
A6_CGTCGGCT_REV	/5Phos/AGCCGACGAGATCGGAAGAGCGGTTTCAGCAGGAATGCCGAG
A7_GACGTCAA_FWD	ACACTCTTCCCTACACGACGCTCTTCCGATCTGACGTCAA*T
A7_GACGTCAA_REV	/5Phos/TTGACGTCAGATCGGAAGAGCGGTTTCAGCAGGAATGCCGAG
A8_GCGTTTCG_FWD	ACACTCTTCCCTACACGACGCTCTTCCGATCTGCGTTTCG*T
A8_GCGTTTCG_REV	/5Phos/CGAAACGCAGATCGGAAGAGCGGTTTCAGCAGGAATGCCGAG

Supplementary Table S4, DNA sequences of the plasmid used for primer extension experiments:

Name	DNA sequence
pBluescript SK(-)-pC3N sequence:	<p>CACCTGACGCGCCCTGTAGCGGCGCATTAAAGCGCGGGTGTGGTGGTTACGCGCAGCGTGACCGG CTACACTTGCCAGCGCCCTAGCGCCCGCTCCTTTTCGCTTTCTTCCCTTCTTTCTCGCCACGTTGCGCG GCTTTCCCGTCAAGCTCTAAATCGGGGGCTCCCTTTAGGGTCCGATTTAGTGCTTTACGGCACCTC GACCCCAAAAACTTGATTAGGGTGTAGTTACGTAGTGGGCCATCGCCCTGATAGACGGTTTTTC GCCCTTTGACGTTGGAGTCCACGTTCTTAATAGTGGACTCTTGTTCAAACTGGAACAACACTCAAC CCTATCTCGGTCTATTCTTTGATTTATAAGGGATTTTGCCGATTTGCGCCTATTGGTTAAAAAATGAG CTGATTTAACAAAAATTAACGCGAATTTAACAAAATATTAACGCTTACAATTTCCATTGCGCATTCA GGCTGCGCAACTGTTGGGAAGGGCGATCGGTGCGGGCCTCTTCGCTATTACGCCAGCTGGCGAAAG GGGGATGTGCTGCAAGGCGATTAAGTTGGTAACGCCAGGGTTTTCCAGTCACGACGTTGTA CGACGGCCAGTGAATTGAATACGACTCACTATAGGGCGAATTGGGTACCGGGCCCCCTCGAGG TCGACGGTATCGATAaGCTTGGGAcccTGGGAGGGAGATCCACTAGTTCTAGAGCGGCCGCCACCGC GGTGGAGCTCCAGCTTTTGTCCCTTTAGTGAGGGTAATTCGAGCTTGGCGTAATCATGGTCATA GCTGTTTCTGTGTGAAATTGTTATCCGCTCACAATCCACACAACATACGAGCCGGAAGCATAAAGT GTAAAGCCTGGGGTGCCTAATGAGTGAGCTAACTCACATTAATTGCGTTGCGCTCACTGCCCGCTTT CCAGTCGGGAAACCTGTCGTGCCAGCTGCATTAATGAATCGGCCAACGCGCGGGGAGAGGCGGTTT GCGTATTGGGCGCTCTCCGCTTCTCGCTCACTGACTCGCTGCGCTCGGTGCTCGGCTGCGGCGA GCGGTATCAGCTCACTCAAAGGCGGTAATACGTTATCCACAGAATCAGGGGATAACGCAGGAAAG AACATGTGAGCAAAGGCCAGCAAAGGCCAGGAACCGTAAAAAGGCCGCTTGTGGCGTTTTTC CATAGGCTCCGCCCCCTGACGAGCATCACAAAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCG ACAGGACTATAAGATACCAGGCGTTTTCCCTGGAAGCTCCCTCGTGCCTCTCTGTTCCGACCT GCCGCTTACCGGATACCTGTCCGCTTTCTCCCTTCGGGAAGCGTGGCGCTTTCTCATAGCTCACGCT GTAGGTATCTCAGTTCGGTGTAGGTGCTTCCGCTCAAGCTGGGCTGTGTGCACGAACCCCCGTTCA GCCGACCGCTGCGCCTTATCCGGTAACTATCGTCTTGAGTCCAACCCGGTAAGACACGACTTATCG CCACTGGCAGCAGCCACTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTC TTGAAGTGGTGGCCTAACTACGGCTACACTAGAAGGACAGTATTTGGTATCTGCGCTCTGCTGAAGC CAGTTACCTTCGGAAAAAGAGTTGGTAGCTCTTGATCCGGCAAACAAACCACCGCTGGTAGCGGTG GTTTTTTGTTTGCAAGCAGCAGATTACGCGCAGAAAAAAGGATCTCAAGAAGATCCTTTGATCTTT TCTACGGGTCTGACGCTCAGTGGAACGAAAACACTCACGTTAAGGGATTTTGGTCATGAGATTATCAA AAAGGATCTTACCTAGATCCTTTAAATTAATAAATGAAGTTTTAAATCAATCTAAAGTATATGAG TAAACTTGGTCTGACAGTTACCAATGCTTAATCAGTGAGGCACCTATCTCAGCGATCTGTCTATTCG TTCATCCATAGTTGCTGACTCCCCGCTGTAGATAACTACGATACGGGAGGGCTTACCATCTGGCC CCAGTGCTGCAATGATACCGCGAGACCCACGCTCACCGCTCCAGATTTATCAGCAATAAACCAGCC AGCCGGAAGGGCCGAGCGCAGAAAGTGGTCTGCAACTTTATCCGCTCCATCCAGTCTATTAATTGT TGCCGGGAAGCTAGAGTAAGTAGTTCGCCAGTTAATAGTTTGCGCAACGTTGTTGCCATTGCTACAG GCATCGTGGTGTACGCTCGTCTTGGTATGGCTTCATTAGCTCCGTTCCCAACGATCAAGGCG AGTTACATGATCCCCATGTTGTGCAAAAAAGCGGTTAGCTCCTTCGGTCTCCGATCGTTGTCAGAA</p>

GTAAGTTGGCCGAGTGTTATCACTCATGGTTATGGCAGCACTGCATAATTCTTACTGTCATGCCA TCCGTAAGATGCTTTTCTGTGACTGGTGAGTACTCAACCAAGTCATTCTGAGAATAGTGTATGCGGC GACCGAGTTGCTCTTGCCCGCGTCAATACGGGATAATACCGCGCCACATAGCAGAACTTTAAAAGT GCTCATCATTGGAAAACGTTCTTCGGGGCGAAAACCTCAAGGATCTTACCGCTGTTGAGATCCAGT TCGATGTAACCCACTCGTGCACCCAAGTATCTTCAGCATCTTTTACTTTCACCAGCGTTTCTGGGTGA GCAAAAACAGGAAGGCAAAATGCCGCAAAAAGGGAATAAGGGCGACACGGAAATGTTGAATACT CATACTCTTCCTTTTCAATATTATTGAAGCATTTATCAGGGTTATTGTCTCATGAGCGGATACATATT TGAATGTATTTAGAAAAATAAACAAATAGGGGTTCCGCGCACATTTCCCCGAAAAGTGC

Supplementary Table S5: Primers used for establishing the *Chaf1a*-dTAG cell line.

	Sequence
sgRNA#1	CGCCGTCGCGGAGATGTTGG AGG
Primer#1	CAATGGCTACTTTCAACCCGTC
Primer#2	CACCCAAACCGACCTTCCTG
Primer#3	GACGTA CTGAGTGACCTCTT
Primer#4	CCAGCCCCTCAATCGTTCAA